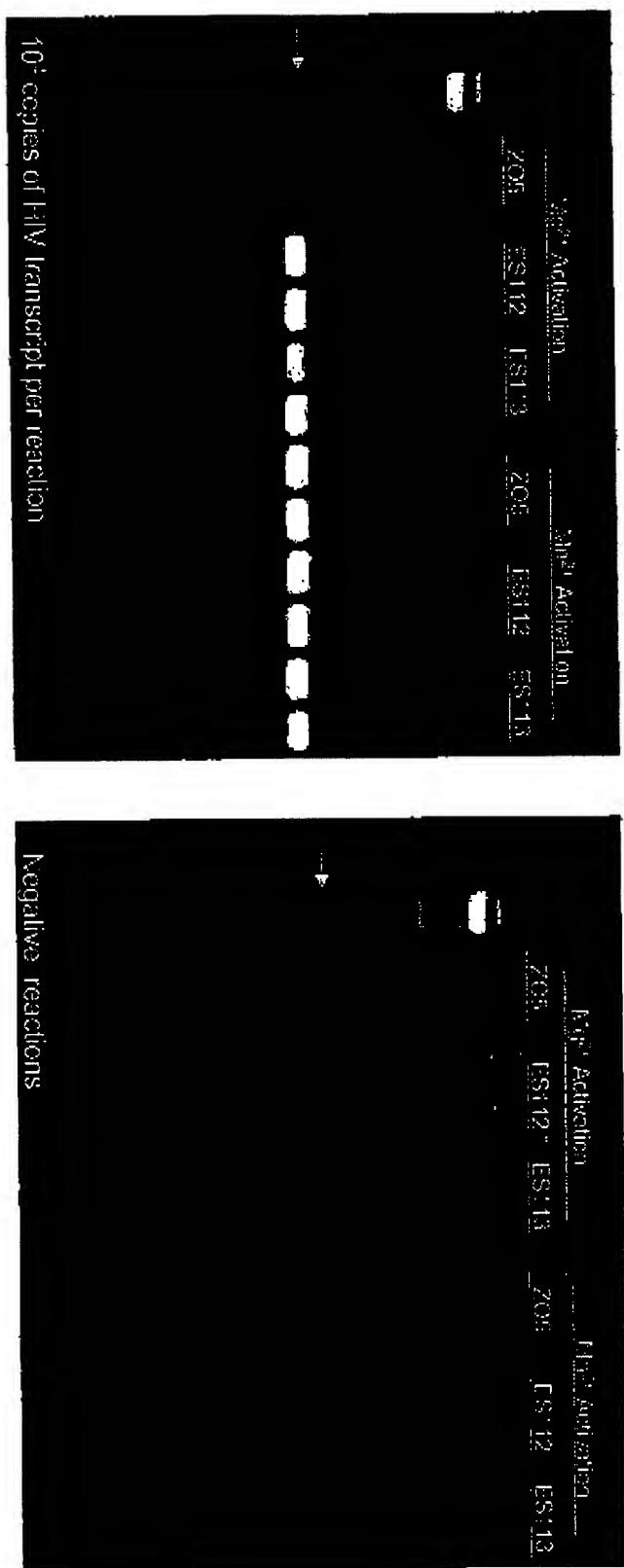


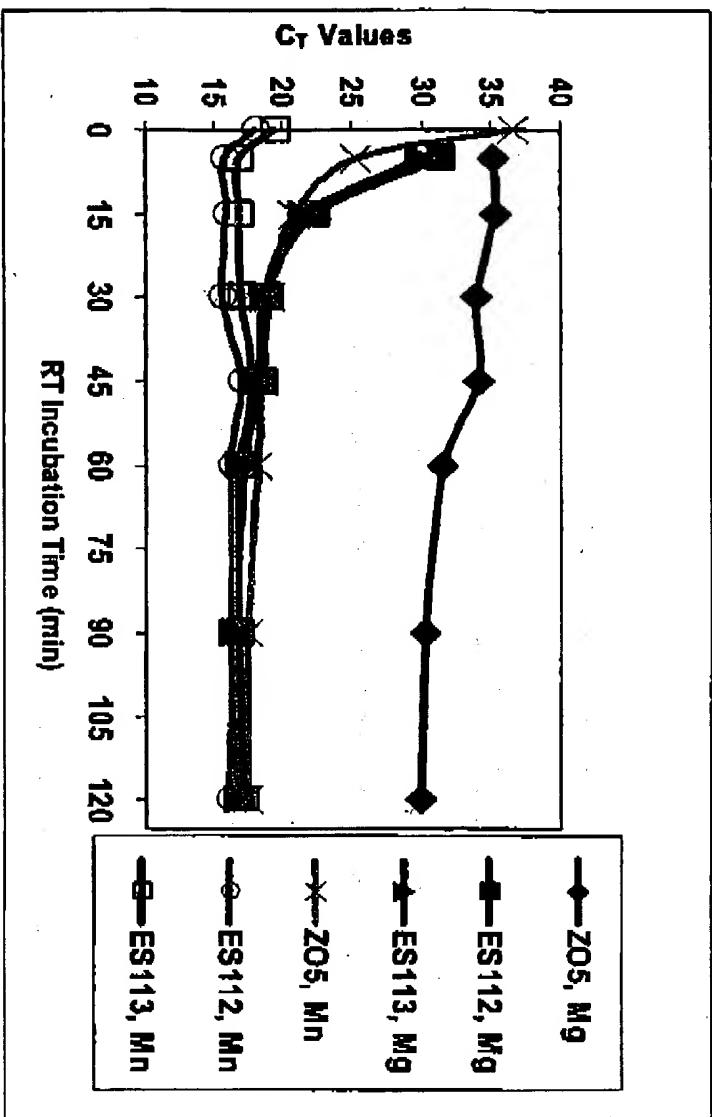
Improved Mg^{2+} -activated RT-PCR with ES112 & ES113



Three different thermostable DNA polymerases were used to reverse transcribe an HIV transcript RNA template and subsequently amplify the cDNA in a coupled RT-PCR in the presence of either 3 mM Mg^{2+} or 3 mM Mn^{2+} . After 55 cycles of PCR, gel results demonstrate specific amplification products from RNA with ZO5 in the presence of Mn^{2+} , but no specific product was observed when Mg^{2+} was used as the divalent metal ion activator. However, designer enzymes ES112 and ES113 produced specific amplification product with either Mg^{2+} or Mn^{2+} activation.

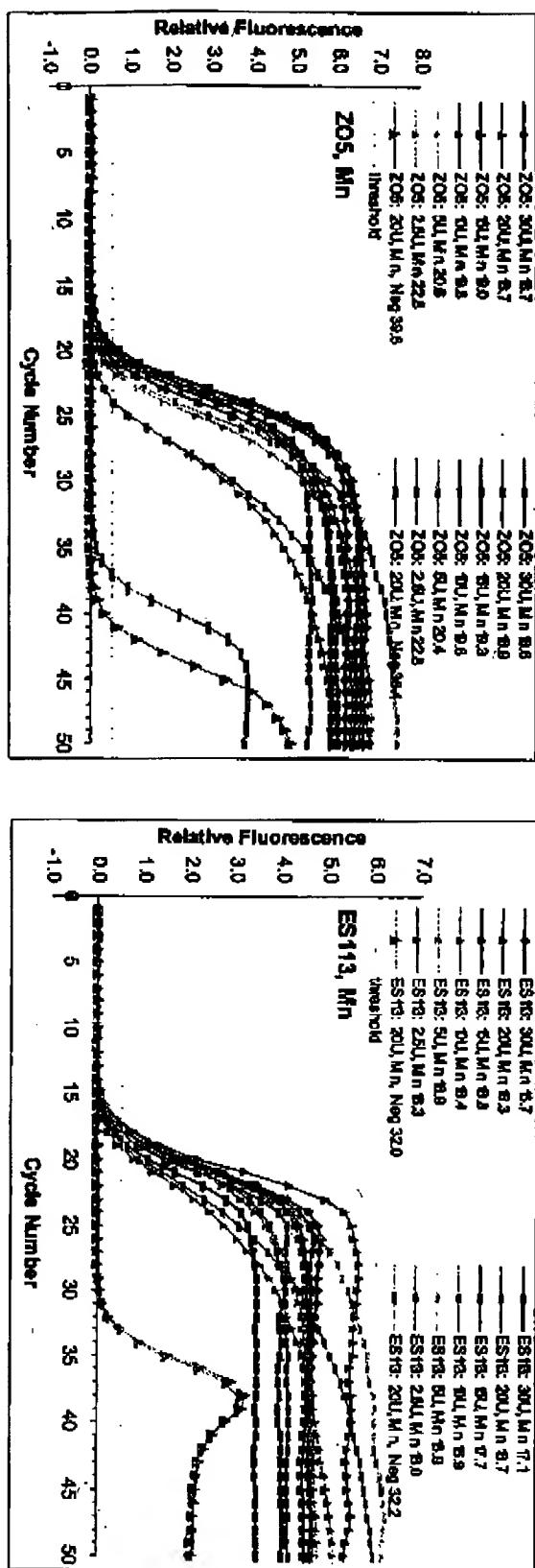
Reduced RT Time Requirement for ES112 & ES113 in Mn^{2+}

A 280 bp GAPDH RNA template was subjected to various RT incubation times and then amplified by PCR. In all cases PCR profiles were identical and the results were analyzed by kinetic PCR. The C_T values of growth curves are plotted in the following chart.



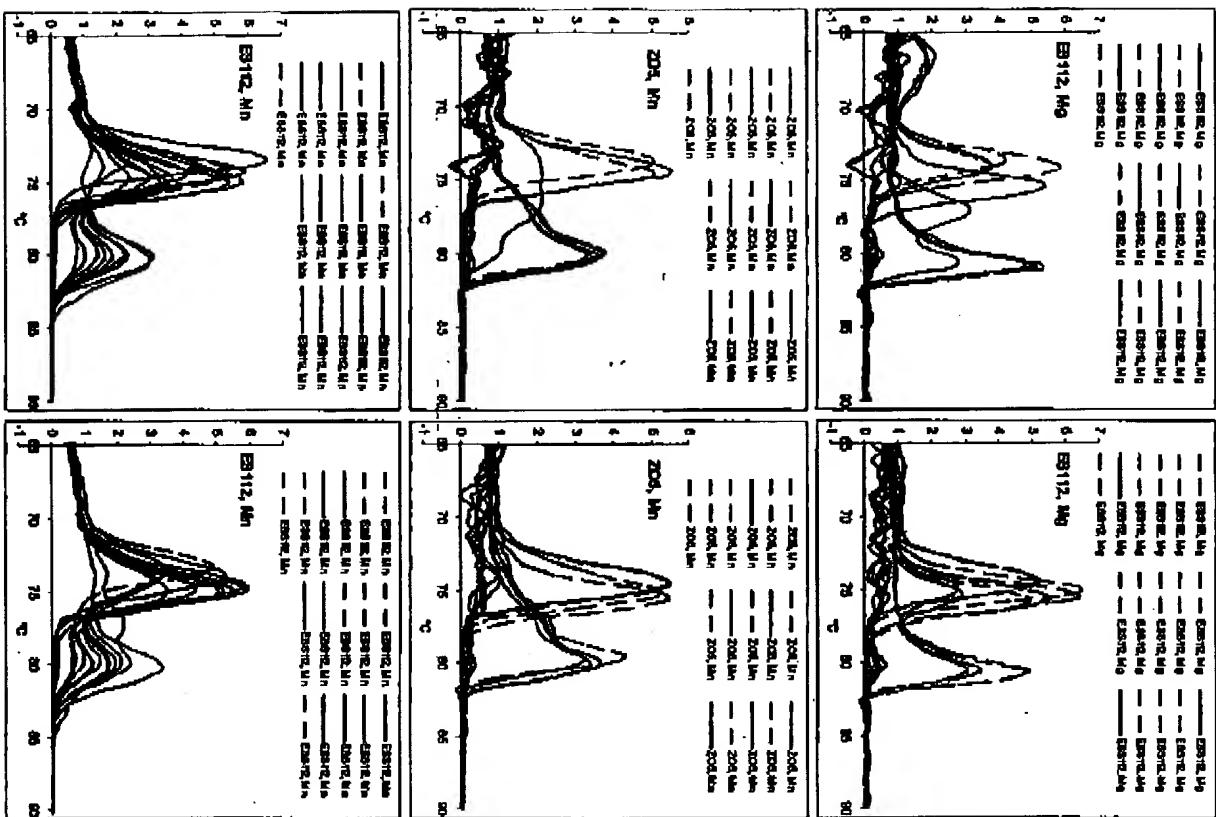
Following a 30 min RT incubation time and Mg^{2+} activation, the mutant enzymes ES112 and ES113 achieved RT activity similar to Mn^{2+} -activated wild-type ZO5 DNA polymerase. With Mn^{2+} activation, the mutant enzymes exhibited similar RT activity, but with much shorter RT incubation times (as low as 5 min). Even with no added RT incubation time there were only slight C_T delays for Mn^{2+} -activated mutant enzyme amplifications and initial PCR ramp times apparently are sufficient for the RT step to occur.

Efficient RT-PCR at Decreased ES112 & ES113 Enzyme Concentrations



Enzyme concentration was titrated from 30 U down to 2.5 U per reaction for Z05, ES112 and ES113. A significantly higher C_T value is observed with 2.5 U of Z05 when compared to higher enzyme concentrations. The ES112 and ES113 perform relatively efficient RT-PCR with as little as 2.5 U of enzyme per 50 μ L reaction.

Improved Low Copy Sensitivity with ES112 in Mn^{2+} -activated RT-PCR



ES112, Mg^{2+}

Nominally 0.5 copies of HIV transcript RNA per reaction

10/32 Positives

were amplified in 50 μ L RT-PCR amplifications optimized for

Mg^{2+} -activated ES112, Mn^{2+} -

activated ES112 or Mn^{2+} -

activated ZO5 ("Gold

Standard"). The T_m of end-point

RT-PCR product was used to

distinguish successful

amplification of transcript RNA

(specific product) from

negative reactions (nonspecific

product). The Mg^{2+} -activated

ES112 reactions had the same

low copy sensitivity as the Mn^{2+} -

activated ZO5, while the low

copy sensitivity was observed to

be twice as good with Mn^{2+} - activated ES112.

ZO5, Mn^{2+}

10/32 Positives

activated ZO5 ("Gold

Standard"). The T_m of end-point

RT-PCR product was used to

distinguish successful

amplification of transcript RNA

(specific product) from

negative reactions (nonspecific

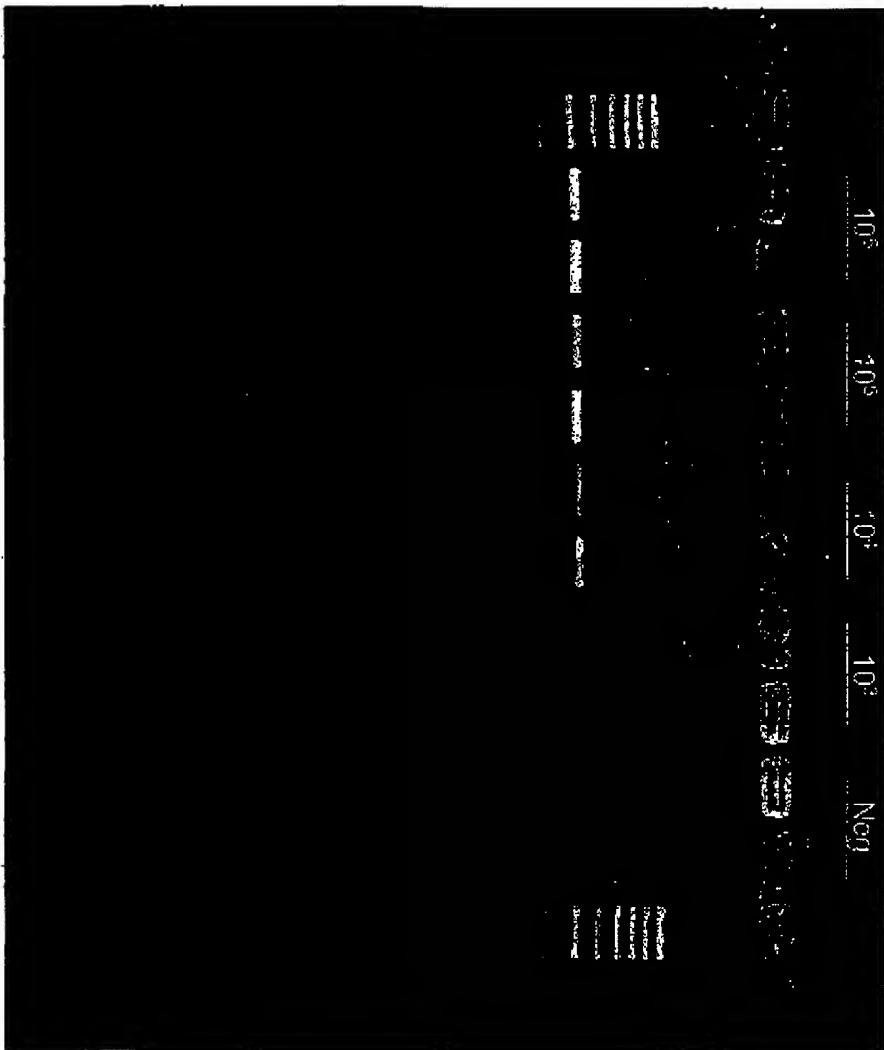
ES112, Mn^{2+}

20/32 Positives

activated ZO5, while the low

copy sensitivity was observed to

be twice as good with Mn^{2+} - activated ES112.



Various concentrations of pAW109 transcript RNA were amplified by single-buffer RT-PCR. All reactions contained 2 mM Mg²⁺ and CS6 DNA polymerase. Following 45 cycles of PCR, products of the correct size were observed with as little as 10³ copies of RNA per reaction. Negative control reactions lacking RNA transcript produced no specific product of the expected amplicon size.